

# Hypoxia-Inducible Factor 1 $\alpha$ Expression in Chorionic Tissue and Decidua of Women with Spontaneous Abortion at the First Trimester of Pregnancy

E.V. Mashkina, K.A. Kovalenko and E.V. Butenko

Southern Federal University, Rostov-on-Don, Russia

## Article history

Received 2014-08-31;

Revised 2014-09-07;

Accepted 2014-10-01

## Corresponding Author:

E.V. Mashkina, Southern Federal University, Rostov-on-Don, Russia

Email: lenmash@mail.ru

**Abstract:** Oxygen-regulated genes expression has important role in pre-implantation embryonic metabolism regulation. Hypoxia Inducible Factor (HIF) regulated by hypoxia oxygen tension is crucial for placenta development. But the data about its role in spontaneous abortion is very poor. Thus, we aimed to determine an expression level of *HIF-1 $\alpha$*  in chorionic tissue and decidua at pregnancy. Samples of chorionic tissue and decidua were taken after surgical termination of normally progressing pregnancies in 5-9 week of gestation (n = 8) and spontaneous abortion in 5-9 week of gestation (n = 9). *HIF-1 $\alpha$*  expression was analyzed using semi-quantitative reverse transcription-polymerase chain reaction. Compared with decidual tissue, the expression of *HIF-1 $\alpha$*  was increased in chorionic tissue in condition of normally progressing pregnancy. *HIF-1 $\alpha$*  expression in samples of both tissues is equal in spontaneous abortion. In same time the expression of *HIF-1 $\alpha$*  was decreased (1,5 fold) in chorionic tissue for spontaneous abortion compared with control group. The results demonstrated that low *HIF-1 $\alpha$*  expression level in chorionic tissue can be associated with spontaneous abortion in first trimester of pregnancy.

**Keywords:** Spontaneous Abortion, HIF-1, Gene Expression, Chorionic Tissue

## Introduction

About 15% of all human pregnancies end in spontaneous abortion before 12 weeks of gestation. The pathophysiology of pregnancy loss is complicated and poorly understood. Major part of the pregnancy loss causes remains unexplained after comprehensive study. Immunity, angiogenesis, apoptosis-related genes are involved in pathogenesis. The aberrant maternal inflammation associated with spontaneous abortion is closely linked to deficient placental perfusion (Renaud *et al.*, 2011).

Early stages of the mammalian placenta development are regulated by oxygen tension and the hypoxic uterine environment (Giaccia *et al.*, 2004). A hypoxic environment is essential for proper embryonic development. Low oxygen appears to prevent trophoblast differentiation into an invasive phenotype. This physiological switch in oxygen tension is a prerequisite for proper placental development (Patel *et al.*, 2010). Low oxygen tension induces embryo development up to the blastocyst stage (Kind *et al.*, 2005; Harvey *et al.*, 2007). Vascular

development during embryonic and fetal growth in utero is triggered by hypoxia (Simon and Keith, 2008).

Oxygen-regulated genes expression plays an important role in pre-implantation embryonic metabolism regulation. Hypoxia Inducible Factor (HIF) regulated by hypoxia oxygen tension is crucial for placenta development. This factor is up-regulated under hypoxic conditions that take place during implantation, fetal placentation, organogenesis, angiogenesis and embryo growth (Adelman *et al.*, 2000). On the other hand, HIF-1 $\alpha$  protein expression can also be induced by other stimuli, for example hormones, cytokines and growth factors (Pringle *et al.*, 2010).

HIF-1 modulates gene transcription by binding to a specific DNA sequence (Hypoxic Response Element (HRE)). HIF-1 is a heterodimer composed of HIF-1 $\alpha$  and HIF-2 $\alpha$  subunits. HIF-1 $\alpha$  and HIF-2 $\alpha$  activate a number of common genes. But HIF-1 $\alpha$  exclusively induces the hypoxic transcription of glycolytic genes such as phosphoglycerate kinase I, aldolase (Wang *et al.*, 2005; Covelto *et al.*, 2006).

HIF is the primary molecular sensor which responds to oxygen tension changes (Adelman *et al.*, 1999; Maltepe *et*

*al.*, 2005). HIF as transcription factor regulates many cellular processes, for example angiogenesis, invasion, erythropoiesis and cell survival (Semenza, 2000; Bruick, 2003; Covello and Simon, 2004; Cowden Dahl *et al.*, 2005a). But the data about its role in spontaneous abortion is very poor.

To further investigate the role of HIF-1 $\alpha$  in spontaneous abortion, we measured the *HIF-1 $\alpha$*  gene expression in chorionic tissue and deciduas.

## Material and Methods

Prior to inclusion in the study, all subjects underwent a standard diagnostic work-up. The women were examined using transvaginal ultrasonography for the absence of uterine abnormalities and polycystic ovary syndrome. Women with previously diagnosed arterial hypertension, diabetes, thyroid diseases, autoimmune pathology and infections during pregnancy were excluded from studied population. Women contacting with exogenous risk factors, such as alcohol, electromagnetic radiation, industrial noise, vibration, chemical pollutants were also excluded. The study was approved by the Southern Federal University Bioethics Committee. The participants willingly signed the informed consent. After approval by institutional review board, 9 women (mean age 29) with spontaneous abortion and 8 women (mean age 29) with normally progressing pregnancies were studied.

Samples of chorionic and decidual tissues were taken after surgical termination by curettage of normally progressing pregnancies in 5-9 week of gestation (n = 8) and spontaneous abortion in 5-9 week of gestation (n = 9). Villous samples from the control group were obtained from women undergoing elective abortion for social reasons. Samples were stored at -80°C in aliquots for RNA isolation and thawed only once to avoid degradation.

Total RNA isolation was extracted by the acid guanidinium thiocyanate phenol method (Chomczynski and Sacchi, 1987). Upon isolation, RNA was immediately treated with DNase I (Syntol, Russia). RNA integrity was assessed using non-denaturing 1,5% agarose gel electrophoresis. Clear 18S and 28S bands were observed with no signs of RNA degradation. The RNA was reverse transcribed immediately following the RNA isolation and the DNase treatment using the "RT kit" (Syntol, Russia) with the template denaturation step and the oligo (dT) primer. Reverse transcription (with M-MLV enzyme) was performed during 50 min incubation at 42°C for 50 minutes, followed by duration of 92°C for 10 min. cDNA samples were stored at -20°C.

Polymerase Chain Reaction (PCR) was performed with commercially available reagents by Syntol (Russia). Sequences of the *HIF-1 $\alpha$* -specific primers were: forward 5'-ATCTCGGCGAAGTAAAGAATCTG-3'; and reverse 5'-GTCACCATCATCTGTGAGAACC-3'. Human  $\beta$ -*Actin* gene was used as a reference gene. Sequences of the  $\beta$ -

*Actin*-specific primers were: 5'-CTTCTACAATGAGCTGGGTG-3'; and 5'-TCATGAGGTAGTCAGTCAGG-3'. PCR was performed according to the protocol for TerCyc thermocycler (DNK Technologiya, Russia). Cycling parameters for *HIF-1 $\alpha$*  were the following: 1 cycle: 94°C for 10 c; 35 cycles: 94°C for 15 c, 64°C for 30 c and 72°C for 30 c; final elongation: 72°C for 2 min.

The PCR products were analyzed by 2% agarose gel electrophoresis. Gel images were captured using GelDoc XR system (Bio-Rad, USA). Densitometry was performed using ImageJ (NIH, USA). The background was subtracted with the rolling ball radius of 50 pixels.

The intensities of the bands of the target gene (*HIF-1 $\alpha$* ) was normalized to that of  $\beta$ -*Actin*. All experiments were conducted in duplicate. Data were analyzed with MedCalc 11.4.2 software using the appropriate non-parametric Mann-Whitney test. P-value <0.05 was considered statistically significant.

## Results

The expression of *HIF-1* differs for chorionic and decidual tissues in condition of normal gestation. Compared with decidual tissue, the expression of *HIF-1 $\alpha$*  was statistically increased in chorionic tissue in condition of normally progressing pregnancy (P = 0.016) (Fig. 1).

*HIF-1 $\alpha$*  expression in samples of both tissues in spontaneous abortion is equal (Fig. 2). Thus the expression of *HIF-1 $\alpha$*  in chorionic tissue in case of spontaneous abortion does not match for normal gestation condition.

There wasn't any difference in the level of *HIF-1 $\alpha$*  expression in decidua in condition of normal pregnancy compared to spontaneous abortion.

Compared with control group, the expression of *HIF-1 $\alpha$*  was decreased (1.5 fold) in chorionic tissue (P = 0.057) in case of spontaneous abortion (Fig. 3).

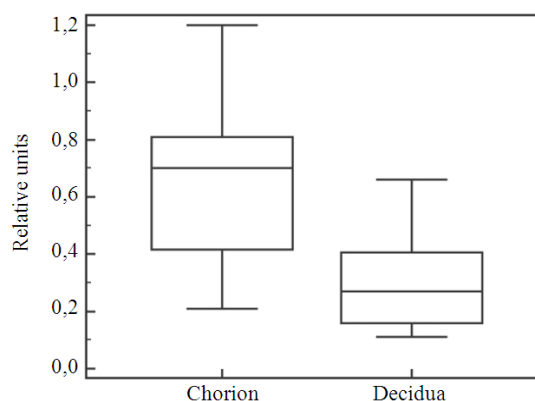


Fig. 1. *HIF-1 $\alpha$*  expression level in chorionic tissue and decidua in condition of normally progressing pregnancy. Gene expression is provided in the same scale in relative units. The mid-lines are medians and the box lines are 25th and 75th percentiles

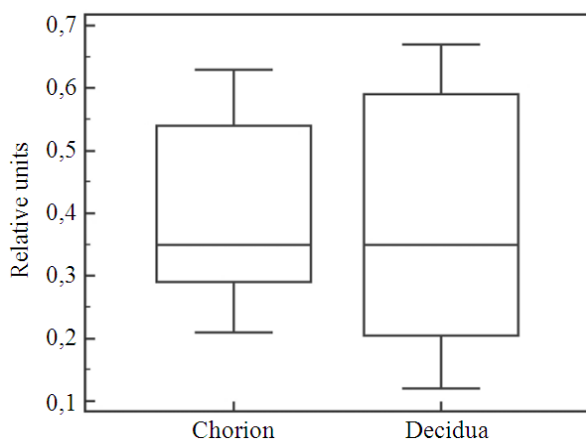


Fig. 2. *HIF-1α* expression level in chorionic tissue and decidua in condition of spontaneous abortion. Gene expression is provided in the same scale in relative units. The mid-lines are medians and the box lines are 25th and 75th percentiles

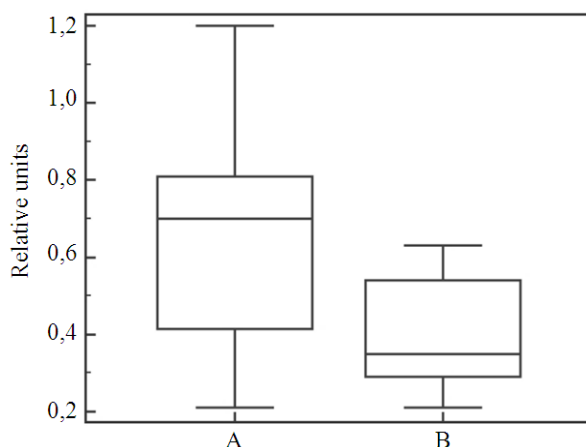


Fig. 3. *HIF-1α* expression level in control group (A) and spontaneous abortion (B) in chorionic tissue. Gene expression is provided in the same scale in relative units. The mid-lines are medians and the box lines are 25th and 75th percentiles

Thus, the high expression level of the *HIF-1α* gene in chorionic tissue is characteristic for a normal pregnancy. The decrease in the expression level of *HIF-1α* gene in chorionic tissue may be associated with miscarriage in the first trimester.

## Discussion

Our study shows, that during normally progressing pregnancy *HIF-1α* expression level in chorionic tissue is significantly increased compared to decidua. In spontaneous abortion *HIF-1α* expression in chorionic tissue decreases and reaches values typical for decidua. This may result in trophoblast differentiation alteration,

implantation changes, or altered angiogenesis in the forming placenta. Furthermore, as a transcription factor, low levels of HIF may have negative effect on organogenesis and embryo growth. On the other hand, low level of *HIF* expression may reflect changes in hypoxic environment and active oxygen radicals level increase, which leads to lipid peroxidation intensification, cell membrane damage and cell death. This statement demands further investigation.

During the first trimester of pregnancy placental oxygen remains low. It appears to be necessary for placental metabolic activity and for protecting placental and fetal tissues against toxic oxygen metabolites (Illsley *et al.*, 2010). The invasion of trophoblast cells is regulated by major different factors including signaling of the adhesion and growth factors regulated by the interactions of decidua and trophoblast (Flaminio and Antczak, 2005; Harris, 2010). Hypoxic conditions are the typical factor that regulates the invasion of trophoblast cells which migrate and invade the surrounding blood vessels of the endometrium in the maternal uterus in persisting hypoxic conditions. Hypoxia induces alteration of various genes including integrin, MMP and TIMP (Luo *et al.*, 2011; Onogi *et al.*, 2011; Na *et al.*, 2012). It was found that the invasion ability of trophoblast is regulated by the expression of *HIF-1α* (Dubinsky *et al.*, 2010). The invasive ability of trophoblast cells decreases according to the inhibition of *HIF-1α* expression by siRNA (Choi *et al.*, 2012).

*HIF-1α* is expressed in syncytiotrophoblast and in villous cytotrophoblast (Rajakumar, 2000). *HIF-1α* mRNA and protein peaked at 7-10 weeks of gestation (Ietta *et al.*, 2006). HIF in hypoxia condition provides a potent stimulus for VEGF synthesis and is essential for development of maternal and placental vasculature in early human pregnancy (Cowden Dahl *et al.*, 2005a; Nau *et al.*, 2002; De Marco and Caniggia, 2002; Daikoku *et al.*, 2003; Qian *et al.*, 2004; Zhang *et al.*, 2009; Arjamaa *et al.*, 2009). *HIF* expression changes exceeding optimal level lead to pathological processes. There is increase level of *HIF* expression in choriocarcinoma and other trophoblastic diseases (Bolat *et al.*, 2010).

Defects in HIF are often responsible for early termination of pregnancy (Goldman-Wohl and Yagel, 2002; Sibai *et al.*, 2005). Complete disruption of HIF signaling results in improper placental development (Fryer and Simon, 2006). Homozygosity for a null allele at the mouse *Hif1a* locus results in embryonic lethality attributable to failed vascularization (Iyer *et al.*, 1998). Cowden Dahl *et al.* (2005b) reported that *HIF-1α*-*HIF-2α* knockout mice displayed a 17% reduction in trophoblast invasion compared with wild type placenta. Several pro- and anti-invasive factors expressed by either the trophoblasts or the decidua were HIF target genes (Cowden Dahl *et al.*, 2005b). These studies

suggest that HIF appear to act as a key mediator in regulation of placental differentiation, growth and function during early pregnancy.

## Conclusion

Our findings show that a low *HIF-1 $\alpha$*  expression level in chorionic tissue (close to values, typical for decidua) can be associated with spontaneous abortion in first trimester of pregnancy.

## Acknowledgement

This study was supported by the federal assignment No. 6.98.2014/K from Russian Ministry of Science and Education. Research was performed with use of the equipment of Collective Using Center of the Southern Federal University "Biotechnology, Biomedicine and Environmental Monitoring" and "High Technology".

We would like to express our appreciation to Rostov-on-Don City Hospital No. 8 for assistance in collecting the clinical material.

## Funding Information

This study was supported by the federal assignment No. 6.98.2014/K from Russian Ministry of Science and Education.

## Author's Contributions

E.V. Mashkina - Conception and design, Acquisition of data, Analysis and interpretation of data, Drafting or revising the article.

K.A. Kovalenko - Analysis and interpretation of data, Contributed unpublished essential data or reagents.

E.V. Butenko - Conception and design, Analysis and interpretation of data, Drafting or revising the article.

## Ethics

We confirm that this manuscript has not been published elsewhere and is not under consideration by another journal. Each author confirms the manuscript represents honest work. All authors have approved the manuscript. Each author agrees with the order in which his name appears on the title page. Study design and methods were approved by Ethics Committee of Southern Federal University.

## References

### 1. Journal paper

Adelman, D., M. Gertsenstein, A. Nagy, M. Simon and E. Maltepe, 2000. Placental cell fates are regulated in vivo by HIF-mediated hypoxia responses. *Genes Dev.*, 14: 3191-3203. DOI: 10.1101/gad.853700

Adelman, D., E. Maltepe and M. Simon, 1999. Multilineage embryonic hematopoiesis requires hypoxic ARNT activity. *Genes Dev.*, 13: 2478-2483.

Arjamaa, O., M. Nikinmaa, A. Salminen and K. Kaarniranta, 2009. Regulatory role of HIF-1 alpha in the pathogenesis of Age-related Macular Degeneration (AMD). *Age. Res. Rev.*, 8: 349-358. DOI: 10.1016/j.arr.2009.06.002

Bolat, F., N. Haberal, N. Tunali, E. Aslan and N. Bal *et al.*, 2010. Expression of Vascular Endothelial Growth Factor (VEGF), Hypoxia Inducible Factor 1 alpha (HIF-1alpha) and Transforming Growth Factors beta1 (TGFbeta1) and beta3 (TGFbeta3) in gestational trophoblastic disease. *Pathol. Res. Pract.*, 206: 19-23. DOI: 10.1016/j.prp.2009.07.017

Bruick, R., 2003. Oxygen sensing in the hypoxic response pathway: Regulation of the hypoxia-inducible transcription factor. *Genes Dev.*, 17: 2614-2623. DOI: 10.1101/gad.1145503

Choi, J., H. Lee, T. Yang and G. Kim, 2012. Effects of hypoxia inducible factors-1 $\alpha$  on autophagy and invasion of trophoblasts. *Clin. Exp. Reproductive Med.*, 39: 73-80. DOI: 10.5653/cerm.2012.39.2.73

Chomczynski, P. and N. Sacchi, 1987. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Analytical Biochem.*, 162: 156-159. DOI: 10.1016/0003-2697(87)90021-2

Covello, K., J. Kehler, H. Yu, J. Gordan and A. Arsham *et al.*, 2006. HIF-2alpha regulates Oct-4: Effects of hypoxia on stem cell function, embryonic development and tumor growth. *Genes Dev.*, 20: 557-570. DOI: 10.1101/gad.1399906

Covello, K. and M. Simon, 2004. HIFs, hypoxia and vascular development. *Current Top. Dev. Biol.*, 62: 37-54. DOI: 10.1016/S0070-2153(04)62002-3

Cowden Dahl, K., B. Fryer, F. Mack, V. Compennolle and E. Maltepe *et al.*, 2005a. Hypoxia-inducible factors 1 $\alpha$  and 2 $\alpha$  regulate trophoblast differentiation. *Molecular Cellular Biol.*, 25: 10479-10491. DOI: 10.1128/MCB.25.23.10479-10491.2005

Cowden Dahl, K., S. Robertson, V. Weaver and M. Simon, 2005b. Hypoxia-inducible factor regulates  $\alpha v \beta 3$  integrin cell surface expression. *Molecular Biol. Cell*, 16: 1901-1912. DOI: 10.1091/mbc.E04-12-1082

Daikoku, T., H. Matsumoto, R. Gupta, S. Das and M. Gassmann *et al.*, 2003. Expression of hypoxia-inducible factors in the peri-implantation mouse uterus is regulated in a cell-specific and ovarian steroid hormone-dependent manner: Evidence for differential function of HIFs during early pregnancy. *J. Biol. Chem.*, 278: 7683-7691. DOI: 10.1074/jbc.M211390200

- De Marco, C. and I. Caniggia, 2002. Mechanisms of oxygen sensing in human trophoblast cells. *Placenta*, 23: S58-S68. DOI: 10.1053/plac.2002.0809
- Dubinsky, V., T. Poehlmann, P. Suman, T. Gentile and U. Markert *et al.*, 2010. Role of regulatory and angiogenic cytokines in invasion of trophoblastic cells. *Am. J. Reproductive Immunol.*, 63: 193-199. DOI: 10.1111/j.1600-0897.2009.00778.x
- Flaminio, M. and D. Antczak, 2005. Inhibition of lymphocyte proliferation and activation: A mechanism used by equine invasive trophoblast to escape the maternal immune response. *Placenta*, 26: 148-159. DOI: 10.1016/j.placenta.2004.05.008
- Fryer, B. and M. Simon, 2006. Hypoxia, HIF and the placenta. *Cell Cycle*, 5: 495-498. DOI: 10.4161/cc.5.5.2497
- Giaccia, A., M. Simon and R. Johnson, 2004. The biology of hypoxia: The role of oxygen sensing in development, normal function and disease. *Genes Dev.*, 18: 2183-2194. DOI: 10.1101/gad.1243304
- Goldman-Wohl, D. and S. Yagel, 2002. Regulation of trophoblast invasion: From normal implantation to preeclampsia. *Molecular Cellular Endocrinol.*, 187: 233-238. DOI: 10.1016/S0303-7207(01)00687-6
- Harris, L., 2010. Review: Trophoblast-vascular cell interactions in early pregnancy: How to remodel a vessel. *Placenta*, 31: S93-S98. DOI: 10.1016/j.placenta.2009.12.012
- Harvey, A., A. Santos, M. Kirstein, K. Kind and B. Fischer *et al.*, 2007. Differential expression of oxygen-regulated genes in bovine blastocysts. *Molecular Reproduct. Dev.*, 74: 290-299. DOI: 10.1002/mrd.20617
- Ietta, F., Y. Wu, J. Winter, J. Xu and J. Wang *et al.*, 2006. Dynamic HIF1 $\alpha$  regulation during human placental development. *Biol. Reproduct.*, 75: 112-121. DOI: 10.1095/biolreprod.106.051557
- Illsley, N., I. Caniggia and S. Zamudio, 2010. Placental metabolic reprogramming: Do changes in the mix of energy-generating substrates modulate fetal growth? *Int. J. Dev. Biol.*, 54: 409-419. DOI: 10.1387/ijdb.082798ni
- Iyer, N., L. Kotch, F. Agani, S. Leung and E. Laughner *et al.*, 1998. Cellular and developmental control of O<sub>2</sub> homeostasis by hypoxia-inducible factor 1 $\alpha$ . *Genes Dev.*, 12: 149-162.
- Kind, K., R. Collett, A. Harvey and J. Thompson, 2005. Oxygen regulated expression of GLUT 1, GLUT 3 and VEGF in the mouse blastocyst. *Molecular Reproduct. Dev.*, 70: 37-44. DOI: 10.1002/mrd.20183
- Luo, J., F. Qiao and X. Yin, 2011. Hypoxia induces FGF2 production by vascular endothelial cells and alters MMP9 and TIMP1 expression in extravillous trophoblasts and their invasiveness in a cocultured model. *J. Reproduct. Dev.*, 57: 84-91. DOI: 10.1262/jrd.10-008K
- Maltepe, E., G. Krampitz, K. Okazaki, K. Red-Horse and W. Mak *et al.*, 2005. Hypoxia-inducible factor-dependent histone deacetylase activity determines stem cell fate in the placenta. *Development*, 132: 3393-3403. DOI: 10.1242/dev.01923
- Na, K., H. Lee, J. Choi, J. Eun and S. Nam *et al.*, 2012. Dynamic alterations in integrin alpha4 expression by hypoxia are involved in trophoblast invasion during early implantation. *J. Cellular Biochem.*, 113: 685-694. DOI: 10.1002/jcb.23398
- Nau, P., T. Van Natta, J. Ralphe, C. Teneyck and K. Bedell *et al.*, 2002. Metabolic adaptation of the fetal and postnatal ovine heart: Regulatory role of hypoxia-inducible factors and nuclear respiratory factor-1. *Pediatric Res.*, 52: 269-278. DOI: 10.1203/00006450-200208000-00021
- Onogi, A., K. Naruse, T. Sado, T. Tsunemi and H. Shigetomi *et al.*, 2011. Hypoxia inhibits invasion of extravillous trophoblast cells through reduction of Matrix Metalloproteinase (MMP)-2 activation in the early first trimester of human pregnancy. *Placenta*, 32: 665-670. DOI: 10.1016/j.placenta.2011.06.023
- Patel, J., K. Landers, R. Mortimer and K. Richard, 2010. Regulation of Hypoxia Inducible Factors (HIF) in hypoxia and normoxia during placental development. *Placenta*, 31: 951-957. DOI: 10.1016/j.placenta.2010.08.008
- Pringle, K., K. Kind, A. Sferruzzi-Perri, J. Thompson and C. Roberts, 2010. Beyond oxygen: Complex regulation and activity of hypoxia inducible factors in pregnancy. *Human Reproduct. Update*, 16: 415-431. DOI: 10.1093/humupd/dmp046
- Qian, D., H. Lin, H. Wang, X. Zhang and D. Liu *et al.*, 2004. Normoxic induction of the hypoxic-inducible factor-1 $\alpha$  by interleukin-1 $\beta$  involves the extracellular signal-regulated kinase 1/2 pathway in normal human cytotrophoblast cells. *Biol. Reproduct.*, 70: 1822-1827. DOI: 10.1095/biolreprod.103.025031
- Rajakumar, A., 2000. Conrad KP. Expression, ontogeny and regulation of hypoxia-inducible transcription factors in the human placenta. *Biol. Reproduct.*, 63: 559-569. DOI: 10.1095/biolreprod63.2.559
- Renaud, S., T. Cotechini, J. Quirt, S. Macdonald-Goodfellow and M. Othman *et al.*, 2011. Spontaneous pregnancy loss mediated by abnormal maternal inflammation in rats is linked to deficient uteroplacental perfusion. *J. Immunol.*, 186: 1799-1808. DOI: 10.4049/jimmunol.1002679

Semenza, G., 2000. HIF-1 and human disease: One highly involved factor. *Genes Dev.*, 14: 1983-1991. DOI: 10.1101/gad.14.16.1983

Sibai, B., G. Dekker and M. Kupferminc. 2005. Pre-eclampsia. *Lancet*, 365: 785-799. DOI: 10.1016/S0140-6736(05)17987-2

Simon, M. and B. Keith, 2008. The role of oxygen availability in embryonic development and stem cell function. *Nat. Rev. Molecular Cell Biol.*, 9: 285-296. DOI: 10.1038/nrm2354

Wang, V., D. Davis, M. Haque, L. Huang and R. Yarchoan, 2005. Differential gene up-regulation by hypoxia-inducible factor-1 $\alpha$  and hypoxia-inducible factor-2 $\alpha$  in HEK293T cells. *Cancer Res.*, 65: 3299-3306. DOI: 10.1158/0008-5472.CAN-04-4130

Zhang, P., X. Zhang, X. Hao, Y. Wang and Y. Hui *et al.*, 2009. Rac1 activates HIF-1 in retinal pigment epithelium cells under hypoxia. *Graefe's Arch. Clin. Exp. Ophthalmol.*, 247: 633-639. DOI: 10.1007/s00417-008-1031-0